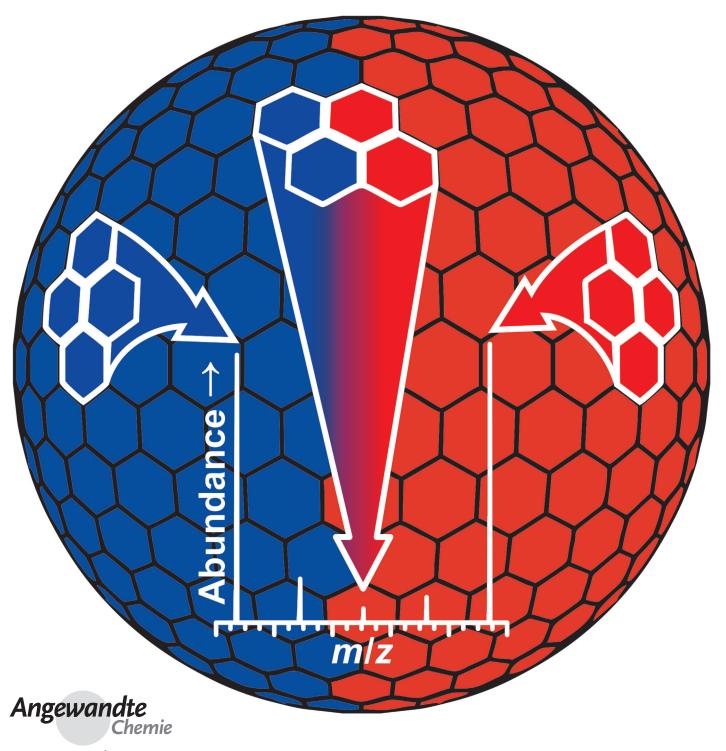




Nanoscale Phase Segregation of Mixed Thiolates on Gold Nanoparticles**

Kellen M. Harkness, Andrzej Balinski, John A. McLean,* and David E. Cliffel*









One of the prized characteristics of the monolayer-protected gold nanoparticle (AuNP) is the versatility of its surface.^[1] The protecting monolayer is comprised of gold-thiolate complexes, the gold-sulfur backbones of which are bound to the gold core while the thiolate tails extend into the surrounding media.^[2] Most molecules containing a thiol group can be integrated into the monolayer, allowing a variety of organic surfaces to be presented on an AuNP scaffold.[1,3] Mixtures of thiols can be utilized to hone chemical functionality and solvation properties, [3a] permitting the exploration of a tremendous amount of chemical space. This chemical space can be further expanded by the formation of ligand domains^[4] through "nanophase separation"^[5] based on ligand-ligand interactions and entropic energy gains.^[6] Surface organization can be harnessed to optimize physical properties and chemical functionalities, from nondestructive membrane transport^[7] to controlled assembly^[8] and ligandabundance-dependent solubility.^[9] In this context, mixedligand AuNPs are somewhat analogous to biomacromolecules, having a versatile nanoarchitecture which can be refined to induce highly specific chemical interactions.^[10]

Existing strategies for characterizing AuNPs with ligand domains, such as scanning tunnelling microscopy^[4a,11] and other spectroscopic techniques,^[4b,12] provide tools limited in their applications for establishing the existence of ligand domains. To advance the development of applications and scientific understanding of nanophase separation in AuNP monolayers, strategies must be developed for their facile and rapid characterization. This methodology must be able to distinguish between AuNP "isomers" with nominally identical molecular formulas and varying molecular structure. This is particularly true for applications targeted to biological systems, where accurate characterization is necessary to understand biological functions.^[13]

We have previously established that gold-thiolate complexes can be desorbed from monolayer-protected AuNP surfaces by the matrix-assisted laser desorption/ionization (MALDI) process, revealing structural characteristics of the protecting monolayer. [14] Ion mobility-mass spectrometry

[*] Dr. K. M. Harkness, A. Balinski, Prof. J. A. McLean, Prof. D. E. Cliffel Department of Chemistry, Vanderbilt University 7330 Stevenson Center, Nashville, TN 37235 (USA) E-mail: john.a.mclean@vanderbilt.edu d.cliffel@vanderbilt.edu

[**] Thanks to Amanda Agrawal, Tracy Okoli, Prof. Brian Huffman, and Dr. Carrie Simpson for providing samples for analysis, Prof. James McBride and Brian Turner for assistance with TEM measurements, Prof. Richard Caprioli for access to instrumentation, and an anonymous reviewer for insightful and helpful comments. Financial support for this work was provided by the Vanderbilt Chemical Biology Interface (CBI) training program (T32 GM065086) and a fellowship of the Vanderbilt Institute of Nanoscale Science and Engineering to K.M.H., the National Institutes of Health (grant numbers GM076479 to D.E.C. and RC2DA028981 for instrumental support to J.A.M.), the Defense Threat Reduction Agency (HDTRA1-09-0013), the Vanderbilt College of Arts and Sciences, the Vanderbilt Institute of Chemical Biology, and the Vanderbilt Institute for Integrative Biosystems Research and Education.

Supporting information for this article is available on the WWW under http://dx.doi.org/10.1002/anie.201102882.

(IM-MS), a two-dimensional gas-phase structural separation technique, is particularly effective for the investigation of desorbed gold-thiolate complexes with high sensitivity. Based on our previous work, we hypothesize that if gold-thiolate complexes are desorbed as discrete portions of the monolayer, nanophase separation in the monolayer will be reflected in the desorbed gold-thiolate ions observed. That is, the relative abundances of homoleptic and heteroleptic gold-thiolate complex ions should reveal the existence and degree of nanophase separation in the monolayer of the parent AuNPs.

To experimentally test our hypothesis and explore the potential of using MALDI-IM-MS for the identification of ligand domains on AuNP surfaces, we synthesized a number of mixed-ligand AuNPs using both one-step mixed-ligand syntheses^[15] and a two-step ligand-exchange process.^[16] The formed AuNPs had core diameter averages between 2-4 nm (see Figure S1 in the Supporting Information). This size range was predicted to have a greater tendency toward complete phase segregation than larger AuNPs. [6,8b] In a typical experiment (Figure 1), these AuNPs were fragmented by MALDI, liberating the gold-thiolate complexes that protect the NP core. The ionized complexes were separated in the gas phase, first by the effective ion surface area and then by the mass-tocharge ratio. A two-dimensional density map was generated, in which dense gold-thiolate ions are clearly separated from organic ions. [14a,b] This gas-phase separation of gold-thiolate complexes from endogenous and exogenous chemical noise allows for the generation of one-dimensional mass spectra which contain only gold-thiolate ions. The Au₄L₄ ion species were selected as the focus of this work because they were the most abundant gold-thiolate ion species observed. [14a-c,17] These Au₄L₄ ion species are either directly desorbed from the AuNP surface^[14a,c] or are products of the rearrangement of "staple" Au_xL_{x+1} species. [2a] In either case, each gold-thiolate complex ion desorbed from the AuNP is expected to contain the ligands present in a given portion of the AuNP surface.

The Au₄L₄ ions within the resulting spectra were identified and their abundances compared to a theoretical model based on the binomial discrete probability distribution. This model, which represents a random distribution of ligands on the AuNP surface, has been used by Dass et al.[18] to study mixed ligand populations on $[Au_{25}L_{18}]^-$ molecules. The binomial distribution can also be used as a theoretical model for mixed ligand populations in gold-thiolate ions liberated from AuNPs. Briefly, the binomial distribution describes the probability of x successes for n binary trials based on the probability (p) of success for any trial. In the context of this work, each trial is considered successful if an alternate ligand, SR', is found, or unsuccessful if an original ligand, SR, is found. The probability of finding an alternate ligand, SR', is determined by its relative abundance on the parent AuNP surface (p is the percentage of SR' in the monolayer).[14b] Each ligand within the Au₄L₄ ion is a single trial, yielding four total trials (n=4), with five possible combinations of SR and SR' within the Au₄L₄ ion species. For a mixed-ligand AuNP with a given composition of SR and SR', the binomial distribution will predict the relative abundances of each Au_4L_4 ion $(Au_4(SR)_4, Au_4(SR)_3(SR')_1...)$.

Communications

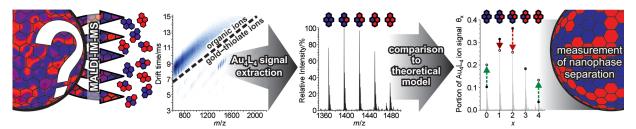


Figure 1. Typical workflow for experiments presented here. Mixed-ligand AuNPs with unknown levels of nanophase separation are analyzed by MALDI-IM-MS. The MALDI process leads to the fragmentation of protecting gold—thiolate complexes from the AuNP surface. The gold—thiolate ions undergo gas-phase separation from organic ions. The Au_4L_4 ion species are extracted from the data by software, and their abundances are compared to a theoretical model based on the binomial distribution. Deviations indicate nanophase separation in the AuNP monolayer.

If the ligands are randomly distributed in the protecting monolayer of the AuNP, the binomial distribution will agree with the Au₄L₄ mass spectral distribution. If nanophase separation is present in the AuNP monolayer, the mass spectral distribution will deviate from the binomial distribution. Homoleptic Au₄(SR)₄ and Au₄(SR)₄ ions will be more abundant than predicted, while heteroleptic Au₄(SR)_x(SR')_{4-x} (1 \geq x \geq 3) ions will be less abundant. Thus the deviation from the binomial distribution can be correlated to the formation of ligand domains.

As an initial control experiment, we created free goldthiolate complexes with a mixture of octanethiol (OT) and [D17]octanethiol ([D17]OT) ligands. These free complexes are not expected to exhibit any significant phase segregation, and the observed ligand distribution agrees well with the binomial model (Figure 2). The residual sum of squares (r), a measurement of deviation from the binomial model, is $2.6 \times$ 10⁻⁴, similar to residual values reported for ligand populations on unfragmented AuNP ions.[18] If the mixed-ligand goldthiolate complexes are generated by place exchange on an AuNP, the residual is generally higher than the control (r is around 10^{-3}). This is true even for ligand mixtures with strong similarities, such as OT:[D17]OT $(r=5.9\times10^{-3})$ or tiopronin:glutathione (Tio:GS, Figure 2). In such cases, the higher residuals may reflect a minimal degree of phase segregation caused by monolayer sites with higher exchange reactivity.^[19] If the ligands differ in chemical functionality or length, as with tiopronin:mercaptoundecanoic acid (Tio:MUA), the residual generally increases to $> 10^{-2}$. If the ligand differences are sufficiently strong, the formation of Janus AuNPs with complete nanophase separation can be observed. These AuNPs, such as the tiopronin:mercaptoundecyltetraethylene glycol (Tio:MUTEG) AuNPs, yield residuals above 10⁻¹. For Janus AuNPs with a 50:50 monolayer composition, the amount of ions containing only SR or SR' should be close to 50% of the total ion signal. This is the case for Tio:MUTEG, where the homoleptic Au₄(Tio)₄ and Au₄-(MUTEG)₄ ion species represent 42 and 45% of the Au₄L₄ ion signal, respectively (Figure 2).

One of the advantages of this strategy is the ability to compare the degree of nanophase separation for multiple AuNP samples. Figure 3 illustrates this ability by comparing the residual sum of squares for various mixed-ligand AuNPs at varying ligand:ligand ratios. In addition to the mixed-ligand

AuNPs described in Figure 2, other ligand:ligand ratios and ligand:ligand combinations were tested: MUA:Tio (beginning with MUA AuNPs and adding tiopronin, the inverse order of Tio:MUA shown in the center of Figure 2), Tio:dPEG4 acid® (a mercaptotetraethylene glycol with a carboxylic acid terminus), Tio:OT, mercaptopropionic (MPA):OT,[4a] nonanethiol:methylbenzenethiol (NT:MBT), [8a,b] OT:decanethiol (DT), [6] and OT:[D17]OT. The lowest residuals are on the order of 10^{-5} , and the pattern illustrated and described in Figure 2 begins to become clear when the residual approaches 10^{-2} . The number of samples scattered between these two values supports the existence of multiple degrees of nanophase separation within a given AuNP sample. [8d,20] Because this technique is averaging with respect to the degree of nanophase separation, the various residuals reflect the relative abundances of AuNPs with phase-segregated or randomly distributed ligands in the monolayer. For most 50:50 binary ligand mixtures obtained by ligand-exchange reactions, some nanophase segregation is observed (Figure 3). This observation is somewhat surprising, given the strong similarities between ligands such as tiopronin and glutathione. No substantial phase segregation was observed for AuNPs generated by mixed-ligand syntheses, even for ligand mixtures with extreme polarity and length differences. Heating MPA:OT AuNPs at 55 °C for 1 h had no apparent effect on the degree of nanophase separation (r=5.7 and 6.1×10^{-3} before and after heating, respectively).

Understanding the lack of phase segregation in mixedligand syntheses may require a closer look at the role of goldthiolate complexes as a precursor to mixed-ligand AuNPs. In solution, the heteroleptic gold-thiolate complexes exhibit no phase segregation (Figure 2). If these complexes are adsorbed to the gold core during the reduction step of the synthesis, the initial state of the AuNP surface will reflect the lack of phase segregation in its component gold-thiolate complexes. Given that nanophase separation requires ligand movement across the AuNP surface post-synthesis, the lack of phase segregation indicates that ligand movement across the surface is minimal. This finding reinforces claims of control over nanophase separation through an interfacial engineering approach, [21] the importance of gold-thiolate complexes to monolayer structure, [14a] and opens the possibility of obtaining diverse AuNP isomers by pursuing multiple synthetic routes.



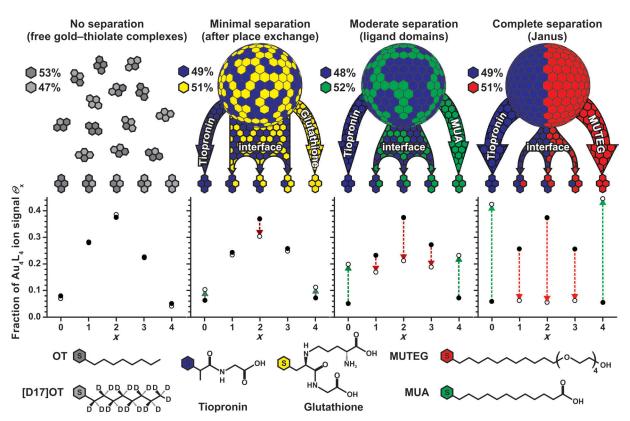


Figure 2. A comparison of experimental and theoretical ligand distributions for free gold—thiolate complexes and three mixed-ligand AuNPs obtained by ligand-exchange reactions of tiopronin AuNPs with free glutathione, 11-mercaptoundecanoic acid (MUA), or mercaptoundecyltetraethylene glycol (MUTEG). Deviation from the theoretical model indicates the presence of phase-segregated gold—thiolate monolayers on AuNPs. Various ligand mixtures yield different degrees of nanophase separation.

In this work we have demonstrated the ability to observe and measure phase segregation in the protecting monolayers of AuNPs by characterizing the gold-thiolate complexes which comprise the monolayer. This ability enables a novel strategy for the analysis of nanophase separations on AuNPs which is rapid, semi-quantitative with respect to the degree of phase segregation, and capable of characterizing a wide variety of ligand mixtures. Using this strategy, we were able to compare mixtures of different ligands, ligand:ligand ratios, and synthetic approaches. We found that nanophase separation is often present, though in varying degrees, and that ligand exchange reactions which combine ligands of varying lengths and minimal ligand-ligand interactions maximize the amount of nanophase separation. Though the latter is not a novel aspect of our findings, the observation serves as a partial validation of the results and points to further insights which will be gained using this technique. The strategy described here functions as an excellent characterization technique for mixed-ligand AuNPs, as well as a helpful starting point for future studies of phase-segregated monolayers on AuNPs.

Experimental Section

Nanoparticle synthesis: Tiopronin- and octanethiol-protected AuNPs were synthesized by one-phase and two-phase methods, respectively, as described elsewhere. [14b] Mercaptoundecanoic acid-protected AuNPs were synthesized using a one-phase approach similar to

tiopronin with the following modifications: the synthesis was performed in methanol at room temperature using a Au/MUA/NaBH₄ ratio of 1:1:10. Some mixed-ligand AuNPs were synthesized using a one-phase approach in ethanol at room temperature. [15b] In each case, Au and the two selected thiols were combined in a 1:0.5:0.5 molar ratio, for a Au/thiol ratio of 1:1 overall. Mixed-ligand gold-thiolate complexes were formed by adding AuCl₄⁻ to a mixture of OT and [D17]OT in chloroform (molar ratio of 1:1.5:1.5). Transmission electron microscopy, thermal gravimetric analysis, and UV/Vis spectroscopy were used in addition to IM-MS for the characterization of the AuNP samples.

Ligand exchange reactions: Homoligand AuNPs were dissolved in around 2 mL of a suitable solvent (deionized water, methanol, or dichloromethane). The AuNPs were then combined with various amounts of the chosen free thiol for up to 72 h to allow equilibration.

Sample preparation: For hydrophobic samples, to a solution of dichloromethane (100 μL) containing AuNPs (roughly 0.5 mg) was added 2-[(2E)-3-(4-tert-butylphenyl)-2-methylprop-2-enylidene]malononitrile (DCTB) matrix (5 mg). A Pasteur pipet was used to deposit roughly 1 μL of the solution on a stainless steel plate. For the remaining samples, a modified sandwich crystallization method^[22] was utilized. A 0.5 μL aliquot of a saturated solution of α -cyano-4-hydroxycinnamic acid (CHCA) matrix was deposited on a stainless steel plate. After drying, a 0.5 μL aliquot of the concentrated sample solution was deposited and dried, followed by another 0.5 μL spot of matrix solution. All spectra were obtained on a Waters Synapt G1 or Synapt G2 HDMS in the positive ion mode. The laser intensity was generally set at 20 % above threshold, the travelling wave velocity was fixed at 300 m s^{-1} while the wave height was ramped from 9 to 16 V. Data were collected for 60 s for each spectrum.

Communications

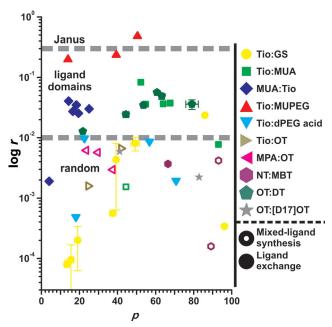


Figure 3. Residual sums of squares (log r, y axis) for various mixed-ligand AuNPs at different ligand:ligand ratios (p, SR:SR', x axis). Specific ligand mixtures are indicated by color and shape; open and filled symbols indicate mixed-ligand syntheses and ligand exchange reactions, respectively. Dashed lines indicate preliminary qualitative assessments of the r values associated with the different types of monolayer structures on AuNPs. The only AuNPs exhibiting phase-segregated monolayers were formed by ligand exchange. Error bars (± 1 standard deviation, n=2) are shown for some Tio:GS and OT:DT AuNPs. Deviations are normally within 3% ligand abundance and 10^{-3} for r.

Data processing and calculations: Mass spectra were extracted from the gold–thiolate region of the ion mobility mass spectrum using Driftscope v2.1 software (Waters Corp.). Peaks were identified and calibrated using MassLynx 4.1 software (Waters Corp.). The processed spectra were exported to Microsoft Excel, which was used for peak identification and isotopic abundance correction as described previously. ^[14b] The peak identification cutoffs were placed at 0.5% abundance relative to the base peak and 10 ppm mass accuracy. All ions with an Au_4L_4 stoichiometry were selected for comparison to a binomial model. For each ligand–ligand combination (i.e., each possible value of x for $Au_4SR_xSR'_{4-x}$), the ion abundances were summed and divided by the total abundance of all Au_4L_4 ions to obtain Θ_x . The binomial distribution was calculated using Microsoft Excel (function "BINOMDIST") with the values n = 4, $0 \le x \le 4$, and Equation (1),

$$p = \sum \frac{xC_x}{n} \tag{1}$$

where C_x is the sum of ion counts for a given value of x.

Received: April 26, 2011 Revised: July 29, 2011

Published online: September 1, 2011

Keywords: mass spectrometry · mixed-valent compounds · monolayers · nanoparticles · self-assembly

- [1] A. C. Templeton, W. P. Wuelfing, R. W. Murray, Acc. Chem. Res. 2000, 33, 27–36.
- a) P. D. Jadzinsky, G. Calero, C. J. Ackerson, D. A. Bushnell, R. D. Kornberg, *Science* 2007, 318, 430–433; b) M. W. Heaven, A. Dass, P. S. White, K. M. Holt, R. W. Murray, J. Am. Chem. Soc. 2008, 130, 3754–3755; c) M. Z. Zhu, W. T. Eckenhoff, T. Pintauer, R. C. Jin, J. Phys. Chem. C 2008, 112, 14221–14224; d) H. Qian, W. T. Eckenhoff, Y. Zhu, T. Pintauer, R. Jin, J. Am. Chem. Soc. 2010, 132, 8280–8281; e) O. Voznyy, J. J. Dubowski, J. T. Yates, P. Maksymovych, J. Am. Chem. Soc. 2009, 131, 12989–12993.
- [3] a) A. C. Templeton, M. J. Hostetler, E. K. Warmoth, S. Chen, C. M. Hartshorn, V. M. Krishnamurthy, M. D. E. Forbes, R. W. Murray, J. Am. Chem. Soc. 1998, 120, 4845–4849; b) M. Brust, M. Walker, D. Bethell, D. J. Schiffrin, R. Whyman, J. Chem. Soc. Chem. Commun. 1994, 801–802; c) A. C. Templeton, S. Chen, S. M. Gross, R. W. Murray, Langmuir 1999, 15, 66–76.
- [4] a) A. M. Jackson, J. W. Myerson, F. Stellacci, *Nat. Mater.* 2004, 3, 330–336; b) C. Gentilini, P. Franchi, E. Mileo, S. Polizzi, M. Lucarini, L. Pasquato, *Angew. Chem.* 2009, 121, 3106–3110; *Angew. Chem. Int. Ed.* 2009, 48, 3060–3064.
- [5] L. H. Radzilowski, S. I. Stupp, Macromolecules 1994, 27, 7747 7753
- [6] C. Singh, P. K. Ghorai, M. A. Horsch, A. M. Jackson, R. G. Larson, F. Stellacci, S. C. Glotzer, *Phys. Rev. Lett.* 2007, 99, 226106.
- [7] A. Verma, O. Uzun, Y. Hu, Y. Hu, H.-S. Han, N. Watson, S. Chen, D. J. Irvine, F. Stellacci, *Nat. Mater.* 2008, 7, 588–595.
- [8] a) G. A. DeVries, M. Brunnbauer, Y. Hu, A. M. Jackson, B. Long, B. T. Neltner, O. Uzun, B. H. Wunsch, F. Stellacci, *Science* 2007, 315, 358-361; b) R. P. Carney, G. A. DeVries, C. Dubois, H. Kim, J. Y. Kim, C. Singh, P. K. Ghorai, J. B. Tracy, R. L. Stiles, R. W. Murray, S. C. Glotzer, F. Stellacci, J. Am. Chem. Soc. 2008, 130, 798-799; c) Q. Xu, X. Kang, R. A. Bogomolni, S. Chen, Langmuir 2010, 26, 14923-14928; d) Y. Hu, O. Uzun, C. Dubois, F. Stellacci, J. Phys. Chem. C 2008, 112, 6279-6284.
- [9] A. Centrone, E. Penzo, M. Sharma, J. W. Myerson, A. M. Jackson, N. Marzari, F. Stellacci, *Proc. Natl. Acad. Sci. USA* 2008, 105, 9886–9891.
- [10] a) A. Hung, S. Mwenifumbo, M. Mager, J. J. Kuna, F. Stellacci, I. Yarovsky, M. M. Stevens, J. Am. Chem. Soc. 2011, 133, 1438–1450; b) X. Liu, Y. Hu, F. Stellacci, Small 2011, Early View.
- [11] A. M. Jackson, Y. Hu, P. J. Silva, F. Stellacci, J. Am. Chem. Soc. 2006, 128, 11135 – 11149.
- [12] a) A. Centrone, Y. Hu, Alicia M. Jackson, G. Zerbi, F. Stellacci, Small 2007, 3, 814–817; b) S. Pradhan, L. Brown, J. Konopelski, S. Chen, J. Nanopart. Res. 2009, 11, 1895–1903.
- [13] D. W. Grainger, D. G. Castner, Adv. Mater. 2008, 20, 867 877.
- [14] a) K. M. Harkness, L. S. Fenn, D. E. Cliffel, J. A. McLean, Anal. Chem. 2010, 82, 3061-3066; b) K. M. Harkness, B. C. Hixson, L. S. Fenn, B. N. Turner, A. C. Rape, C. A. Simpson, B. J. Huffman, T. C. Okoli, J. A. McLean, D. E. Cliffel, Anal. Chem. 2010, 82, 9268-9274; c) A. P. Gies, D. M. Hercules, A. E. Gerdon, D. E. Cliffel, J. Am. Chem. Soc. 2007, 129, 1095-1104; d) K. M. Harkness, D. E. Cliffel, J. A. McLean, Analyst 2010, 135, 868-874.
- [15] a) S. Chen, R. W. Murray, J. Phys. Chem. B 1999, 103, 9996–10000; b) H. Choo, E. Cutler, Y.-S. Shon, Langmuir 2003, 19, 8555–8559.
- [16] R. S. Ingram, M. J. Hostetler, R. W. Murray, J. Am. Chem. Soc. 1997, 119, 9175 – 9178.
- [17] a) C. A. Fields-Zinna, J. S. Sampson, M. C. Crowe, J. B. Tracy, J. F. Parker, A. M. deNey, D. C. Muddiman, R. W. Murray, J. Am. Chem. Soc. 2009, 131, 13844–13851; b) C. A. Fields-Zinna, R. Sardar, C. A. Beasley, R. W. Murray, J. Am. Chem. Soc. 2009,



- 131, 16266–16271; c) Z. Tang, B. Xu, B. Wu, M. W. Germann, G. Wang, J. Am. Chem. Soc. **2010**, 132, 3367–3374.
- [18] A. Dass, K. Holt, J. F. Parker, S. W. Feldberg, R. W. Murray, J. Phys. Chem. C 2008, 112, 20276 – 20283.
- [19] M. J. Hostetler, A. C. Templeton, R. W. Murray, *Langmuir* 1999, 15, 3782–3789.
- [20] Y. Hu, B. H. Wunsch, S. Sahni, F. Stellacci, J. Scanning Probe Microsc. 2009, 4, 24–35.
- [21] a) S. Pradhan, L. Xu, S. Chen, Adv. Funct. Mater. 2007, 17, 2385 –
 2392; b) J. J. Kuna, K. Voitchovsky, C. Singh, H. Jiang, S. Mwenifumbo, P. K. Ghorai, M. M. Stevens, S. C. Glotzer, F. Stellacci, Nat. Mater. 2009, 8, 837 842.
- [22] L. Li, R. E. Golding, R. M. Whittal, J. Am. Chem. Soc. **1996**, 118, 11662–11663.